

# Effects of Salicylic Acid Treatments on Seedling Growth of Red Cabbage under Salt Stress

Chaohong Li

Department of Horticulture, University of Science and Technology Liao Ning, Anshan, China

**Abstract:** High soil salinity is a limiting factor to red cabbage development and production. A study was conducted to investigate the effects of salicylic acid (SA) treatments (0.2 mg plant<sup>-1</sup>) on the development of red cabbage seedlings grown under salt stress (200 mM NaCl). Seedling growth parameters and plant nutrient contents were measured. Salicylic acid treatments had positive effects on plant fresh biomass, dry biomass and root lengths. The plant fresh biomass value of 244.5 mg plant<sup>-1</sup> under the control (zero application of SA) increased to 309.5 mg plant<sup>-1</sup> with SA treatments. The fresh biomass of 205.0 mg plant<sup>-1</sup> under saline conditions increased to 300.0 mg plant<sup>-1</sup> with SA treatments. Similar, plant dry biomass values increased with SA treatments. Furthermore, SA treatments increased root length from 9.21 cm under salt stress conditions (200 mM) to 11.69 cm. In addition to improving plant growth parameters, SA treatments significantly reduced plant Na levels and significantly increased plant contents of K, Ca, P, Fe, Zn and B. SA treatments increased K/Na ratio (a significant indicator of plant tolerance to salinity) from 0.43 to 0.64 and Ca/Na ratio from 0.24 to 0.34.

**Keywords:** *Brassica oleracea*, salicylic acid, dry biomass, root length

## 1. Introduction

Stress factors, especially soil salinity, inhibit or limit plant growth and development. Turkey is among the important agricultural countries of the world. However, about 1.5 million ha land area of Turkey is under the threat of soil salinity [1]. High soil salinity levels are toxic to plants and seriously hinder plant growth and development [2]. High soil salinity levels also reduce leaf sizes, thus reduce photosynthetic activity [3] and reduce enzyme activity and internal hormone production in plants [4]. Cabbages (*Brassica oleracea* L.) are moderately sensitive to saline conditions [5, 6], but there are great varietal variations in tolerance to salt, within the same cultivars and even among ecotypes of a variety [7, 8]. Reclamation of saline soils is both an expensive and a time-consuming process. Therefore, growing a plant on such problematic soils is possible only with certain practices or implementations. Improving soil organic matter content [9], applications of methyl jasmonate [10], 24-Epibrassinolid [11] and salicylic acid [12] like chemicals are the most common methods applied in plant culture over saline soils. Salicylic acid (SA) and similar hormones influence various physiological and biochemical functions of the plants and reduce or inhibit the negative impacts of biotic and abiotic stressors on plants [13, 14]. SA plays a regulatory role in plant tolerance to stress conditions [15-20]. The significance of salicylic acid under biotic and abiotic stress conditions and the role of salicylic acid in transcriptional regulation of defense genes of the plants against such stress conditions were comprehensively investigated and assessed by previous researchers [21]. Salicylic acid applications have been reported to significantly reduce the damage caused by salt stress in vegetable crops such as tomato [22], pepper [23], eggplant [24], sweet basil [25] and faba bean [26]. Red cabbages are widely grown in Turkey. They are quite rich in antioxidants and fibers and thus play a significant role in human diets as a winter vegetable. As it was in various other countries of the world, red cabbage has a great economic significance also in

Turkey. Annual red cabbage production of Turkey was 186.826 tons in 2016 [27].

The present study was conducted to investigate the effects of salicylic acid treatments on plant growth and nutrient contents of red cabbage seedlings under salt stress.

## 2. Material and Methods

Mohrenkkopf (Berk&Han) red cabbage cultivar was used as the plant material for the present study. The seeds were sown into 45-cell 5 x 9 rows viols filled with peat soil. 2-3 seeds were sown per cell. Following the homogeneous emergence, the number of plant per cell was thinned to 1 seedling. In present experiments, 0 mM (control) and 200 mM NaCl (Tekkim, TK.170540.01000, Bursa, Turkey) salt doses and 0 mg plant<sup>-1</sup> (control) and 0.2 mg/plant salicylic acid (SIGMA, 27301, St. Louis, Missouri, USA) doses were applied. Salicylic acid was prepared by diluting to the 0.04 mg ml<sup>-1</sup> from the stock solution (1 mg ml<sup>-1</sup>). Five ml of diluted (0.04 mg ml<sup>-1</sup>) salicylic acid was applied to each of the plant root regions when seedlings had their first true leaves. Following salicylic acid treatments, 5 days later, plants were subjected to salt stress. To prevent acute damages, 200 mM salt treatment was applied in 2-day intervals in 2 portions (firstly 100 mM and 2 days after other 100 mM). The same amount of distilled water was applied to control treatments. Experiments were conducted in randomized plots design with 3 replications and 45 plants in each replicate. Seedlings were left for normal growth and cultural practices were performed whenever required. The vegetative parameters were measured including (i) the fresh biomass, (ii) the dry weight and (iii) the root length. Nutrient contents were also determined. The fresh biomass per plant was obtained by dividing the fresh weight of the plants cut from the upper portion of the roots and weighted with a digital scale ( $e = \pm 0.001$  g) by the total number of plants. Fresh weighted plants were then dried at 72°C for 48 hours in an oven and weighted again with a digital scale ( $e = \pm 0.001$  g). Dry weight was divided by the number of plants to get dry biomass per plant

expressed in  $\text{mg plant}^{-1}$ . Root lengths (cm) were measured with a ruler. Measurement of seedlings contents in sodium (Na), potassium (K), calcium (Ca), magnesium (Mg), phosphorus (P), iron (Fe), zinc (Zn), manganese (Mn), boron (B) and copper (Cu) were performed by an inductively coupled plasma optical emission spectrometer (ICP-OES, Vista-Pro Axial, Agilent Tech., Mulgrave, AUSTRALIA). Prior to the measurement, 0.2 g of dry leaf sample was digested in a closed microwave digestion system (Marsexpress Cem Corp., Matthews, NC, USA) in the presence of 5 ml of concentrated nitric acid ( $\text{HNO}_3$ ) and 2 ml of hydrogen peroxide ( $\text{H}_2\text{O}_2$ ). Deionized water ( $\text{H}_2\text{O}$ ) was used to top-up the volume of the samples to 20 ml. The analytical data was compared to the certified values of a standard reference material (SRM 1573a Tomato Leaf, National Institute of Standards and Technology, Gaithersburg, MD, USA). The data obtained were subjected to an analysis of variance using JMP v.10. statistical software (SAS Institute Inc., USA). Mean values of the parameters measured were compared using LSD test at 5% probability.

### 3. Results and Discussion

Results on the fresh biomass, dry biomass and root length values of red cabbage seedlings grown under salt stress are given in Table 1.

**Table 1:** Effects of salicylic acid on fresh biomass, dry biomass and root length of cabbage seedlings grown under salt stress

NaCl mM	SA $\text{mg plant}^{-1}$	Fw $\text{mg plant}^{-1}$	Dw $\text{mg plant}^{-1}$	RI $\text{mg plant}^{-1}$
0	0	244.5 c	11.65 b	11.65 b
	0.2	309.5 a	16.81 a	16.81 a
200	0	205.0 d	9.21 c	9.21 c
	0.2	300.0	11.69 b	11.69 b
Means of Treatments				
NaCl	0	277.0 A	45.0 A	14.23 A
	200	252.5 B	35.4 B	10.45 B
SA	0	224.8 B	34.9 B	10.43 B
	0.2	304.8 A	45.4 A	14.25 A
LSD	NaCl x SA	3.45	1.51	0.97
	NaCl	2.44	1.07	0.68
	SA	2.44	1.07	0.68

LSD: Least significant difference; means followed by same letter are not statistically different; Fw: fresh biomass; Dw: dry biomass; RI: root length

NaCl x SA interactions had significant effects on fresh biomass, dry biomass and root length of the plants. In fact, the fresh biomass, dry biomass and the root length were lowered by NaCl. SA treatment rose them, as clearly shown in Table 1. The greatest fresh biomass ( $309.5 \text{ mg plant}^{-1}$ ) was obtained from 0 mM NaCl + 0.2 mg SA treatments and the lowest fresh biomass ( $205.0 \text{ mg plant}^{-1}$ ) was obtained from 200 mM NaCl + 0 mg SA treatments. A similar case was also observed for dry biomass values of the plants. The highest dry biomass ( $52.1 \text{ mg plant}^{-1}$ ) was obtained from 0 mM NaCl + 0.2 mg SA treatments and the lowest dry biomass ( $31.9 \text{ mg plant}^{-1}$ ) was obtained from 200 mM NaCl + 0 mg SA treatments. When compared to the control, NaCl treatment reduced fresh and dry biomass of the seedlings. Salt stress-induced reduction of plant weight and limitations on plant

growth and development were also reported for various plants by previous researchers [28-35]. Compared to its control SA treatments significantly increased fresh and dry biomass values. Significant roles of SA in plant growth and development were also reported in previous studies [36, 37]. Dry biomass values of NaCl x SA treatments were similar with the values under control conditions. As it was reported for wheat [38], maize [16], paddy [39] and tomato [40] present findings also revealed that salt stress-induced regress in plant growth could be reduced by applications of SA. SA treatments also increased root length from 9.21 cm under salt stress to 11.69 cm. Similar improved rooting with SA treatments were also reported by previous researchers [41-44].

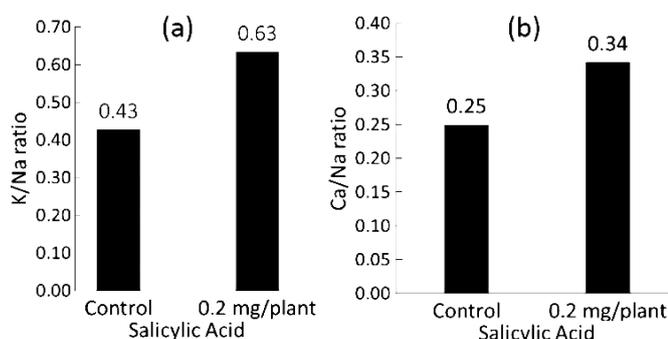
Data on Na, K, Ca, Mg, P, Fe, Zn, Mn, B and Cu contents of red cabbage seedlings grown under salt stress are presented in Table 2. Compared to the control NaCl treatment clearly increased Na contents. Such increases in Na contents with NaCl treatments were also commonly reported by previous studies [29, 32, 45, 46]. With regard to other macro and micro nutrients, it was observed that NaCl x SA interaction had significant effects on K, Ca, P, Fe, Zn and B contents of red cabbage seedlings (Table 2). NaCl treatments led to insignificant increases in K and Ca contents, but significant decreases in P and B contents. The interaction between soil salinity and P content is quite complex phenomenon and plant P respond to saline conditions (increase, decrease or insignificant change) vary based on plant species [47]. In present the study, the decrease in P contents under saline conditions was significantly tempered by SA treatments and even an increase was achieved in P contents when compared to the control (zero SA). NaCl treatments reduced plant boron contents by about 40% ( $0.58 \mu\text{g plant}^{-1}$  in control and  $0.35 \mu\text{g plant}^{-1}$  under saline condition). The decrease in green herbage boron concentrations with salinity may be related to reduced transpiration flow hindering passive transport of boron [48]. Increase or decrease in plant boron contents under saline conditions are also related to genotypic differences, plant age, climatic conditions, and especially to the analyzed section of the plants [49]. Reduced salt stress-induced decreases in boron uptake with SA treatments and similar boron concentrations of NaCl x SA treatments to those of the control conditions reflect the positive impacts of salicylic acid on plant growth and development. As compared to NaCl treatments, NaCl x SA treatments significantly increased plant K, Ca, P, Fe, Zn and B contents. Considerable increases were also observed in other elements, but those increases were not found to be significant. Ahmed et al. [50], reported that SA treatment increased the K and B contents of tomato plants in saline condition. Similar findings were also reported for barley [15], maize [16] and paddy [39]. When compared only to NaCl treatments, NaCl x SA treatments significantly increased Zn (from  $0.83$  to  $1.27 \mu\text{g plant}^{-1}$ ) and B (from  $0.35$  to  $0.57 \mu\text{g plant}^{-1}$ ) contents of the plants. Such increases revealed the significant roles of these elements in preservation of root cell membranes through SA. Thusly, Zn was reported to control integrity and permeability of root cell membranes [51, 52]. Similarly, boron was reported to play a significant role in both structural and functional integrity of bio-membranes, especially of plasma bio-membranes [53].

**Table 2:** Effect of salicylic acid on nutritional content of red cabbage seedling grown in saline condition

NaCl mM	SA mg plant <sup>-1</sup>	Na mg plant <sup>-1</sup>	K mg plant <sup>-1</sup>	Ca mg plant <sup>-1</sup>	Mg mg plant <sup>-1</sup>	P mg plant <sup>-1</sup>	Fe µg plant <sup>-1</sup>	Zn µg plant <sup>-1</sup>	Mn µg plant <sup>-1</sup>	B µg plant <sup>-1</sup>	Cu µg plant <sup>-1</sup>
0	0	0.08c	0.96c	0.56c	0.09	0.17c	0.67c	0.89b	0.46	0.58b	0.11
	0.2	0.05c	1.54a	0.86a	0.12	0.28a	1.10a	1.23a	0.64	0.89a	0.16
200	0	2.36a	1.01c	0.59c	0.08	0.14d	0.70c	0.83c	0.82	0.35c	0.11
	0.2	2.16b	1.37b	0.74b	0.11	0.20b	0.97b	1.27a	0.95	0.57b	0.15
Means of Treatments											
NaCl mM	0	0.06B	1.25A	0.71A	0.11A	0.22A	0.88A	1.06	0.55B	0.74A	0.13
	200	2.26A	1.19B	0.66B	0.09B	0.16B	0.83B	1.05	0.89A	0.46B	0.13
SA mg plant <sup>-1</sup>	0	1.22A	0.98B	0.58B	0.09B	0.15B	0.68B	0.86B	0.64B	0.47B	0.11A
	0.2	1.10B	1.45A	0.80A	0.12A	0.24A	1.03A	1.25A	0.79A	0.73A	0.15B
LSD 5%	NaCl x SA	0.11	0.10	0.059	ns	0.013	0.076	0.048	ns	0.06	ns
	NaCl	0.049	0.035	0.024	0.012	0.007	0.031	ns	0.046	0.027	ns
	SA	0.049	0.035	0.024	0.012	0.007	0.031	0.033	0.046	0.027	0.007

LSD- Least significant differences; ns- not significant

Salicylic acid treatments resulted in significant decreases in Na content of seedlings under both control and saline conditions and led to significant increases in the other nutrient contents. While the average Na content of the plants without SA treatments was 1.22 mg plant<sup>-1</sup>, average Na content of SA-treated plants was 1.10 mg plant<sup>-1</sup> (Table 2). Such a case can be explained as plant growth-induced dilution with SA treatments. Similar findings were also reported in wheat [54]. Salicylic acid has various physiological roles in plants including growth, photosynthesis and nutrient uptake [55]. It was reported in the literature that plant nutrient solutions in soil got into the plants through the roots and a healthy and strong root system together with some other factors (SA application in present study) played a significant role in plant growth and development [56]. Compared to NaCl treatments, NaCl x SA treatments reduced Na levels, but increased K and Ca levels, thus increased K/Na ratio from 0.42 to 0.63 and Ca/Na ratio from 0.25 to 0.34 in saline conditions (Figure 1a and b).

**Figure 1:** Effects of salicylic acid on K/Na (a) and Ca/Na (b) ratios of red cabbage seedlings under saline conditions

High K/Na and Ca/Na ratios were significant indicators of tolerance of plants to salt. Ashraf and Khanum [57] showed that K/Na and Ca/Na ratio were higher in salt tolerant sipping wheat line S24 than salt sensitive line Yecora Rojo. Zeng et al., [58] reported that the salt-tolerant rice lines led to the higher K/Na ratio than sensitive ones. Daşgan et al., [45, 59] pointed out that the higher K/Na and Ca/Na ratio the less salt injury in tomato and beans. Plant potassium uptake through active absorption and accumulation increase osmotic pressure and more water influent is provided to the plants. Therefore, potassium plays a significant role in preservation of plant water balance. Salt tolerant species can transport greater

quantities of potassium from the roots to green herbage and hinder plant Na uptakes [45, 60, 61]. Higher Ca/Na ratios of the present study obtained through salicylic acid treatments under saline conditions was provided by prevention of disintegration in cell membrane permeability. Similarly, Tufail et al., [62] reported that SA increased both K/Na and Ca/Na ratio in maize in saline conditions.

#### 4. Conclusions

Salt stress affected negatively the plant growth by means of decreasing some morphological parameters and changing plant nutrients' content in red cabbage. SA treatments had positive effects on growth of red cabbage seedlings under salt stress. The ameliorative effect of SA could be evaluated by the fact that it promotes plant's uptake K or Ca instead of Na in salt stress environment. In comparison to saline conditions (only NaCl treatments), NaCl x SA treatments significantly increased plant K and Ca contents and decreased Na contents. Subsequently, there was an increase of K/Na and Ca/Na ratios of the plants and hence an increase of red cabbage's tolerance to salt stress.

#### References

- [1] Ekmekçi E, Apan M, Kara T. Tuzluluğun bitki gelişimine etkisi. *OMU, Ziraat Fakültesi Dergisi* 2005, **20(3)**: 118-125.
- [2] Saqib ZA, Akhtar J, Haq M, Ahmad I. Salt induced changes in leaf phenology of wheat plants are regulated by accumulation and distribution pattern of Na ion. *Pak. J. Agric. Sci.* 2012, **49**: 141-148.
- [3] Ashraf M, Harris P. Photosynthesis under stressful environments: an overview. *Photosynthetica* 2013; **51(2)**: 163-190.
- [4] Apel K, Hirt H. Reactive oxygen species: metabolism, oxidative stress, and signal transduction. *Annu. Rev. Plant Biol.* 2004; **55**: 373-399.
- [5] Bernstein L, Ayers A. Salt tolerance of cabbage and broccoli, Riverside, CA, 1949; 39 pp.
- [6] Osawa T. Studies on the salt tolerance of vegetable crops in sand culture. *II. Journal of the Japanese Society for Horticultural Science* 1961; **30(1)**: 48-56.

- [7] Cszinszky, A., Optimum planting time, plant spacing, and nitrogen and potassium rates to maximize yield of green cauliflower. *HortScience* 1996, 31 (6), 930-933.
- [8] Maas EV, Grattan S. Crop yields as affected by salinity. *Agronomy* 1999; **38**: 55-110.
- [9] Ekbiç E, Keskin A. Tuz stresi koşullarında yetiştirilen soğanda çay atığı kompostu uygulamalarının etkileri. *Akademik Ziraat Dergisi* 2018; **7(1)**: 1-8.
- [10] Sadeghipour O. Amelioration of salinity tolerance in cowpea plants by seed treatment with methyl jasmonate. *Legume Research: An International Journal* 2017; **40(6)**: 1100-1106.
- [11] Yılmaz Gökdoğan E, Bürün B. 24-Epibrassinolid ön uygulaması yapılmış domates (*Lycopersicon esculentum* Mill.) tohumlarının NaCl stresi koşullarında çimlenmesi ve fide gelişimi. *Afyon Kocatepe Üniversitesi Fen ve Mühendislik Bilimleri Dergisi* 2015; **15(3)**: 18-27.
- [12] Khulenjani MB, Salamati MS. The effects of salinity and methyl jasmonate on some morphological traits of chamomile (*Matricaria chamomilla*). *Research on Crop Ecophysiology* 2015; **10(2)**: 41-49.
- [13] Arfan M, Athar HR, Ashraf M. Does exogenous application of salicylic acid through the rooting medium modulate growth and photosynthetic capacity in two differently adapted spring wheat cultivars under salt stress? *Journal of Plant Physiology* 2007; **164(6)**: 685-694.
- [14] Van Breusegem F, Vranová E, Dat JF, Inzé D. The role of active oxygen species in plant signal transduction. *Plant Science* 2001; **161(3)**: 405-414.
- [15] El-Tayeb M. Response of barley grains to the interactive effect of salinity. *Plant Growth Regulation* 2005; **45(3)**: 215-224.
- [16] Gunes A, Inal A, Alpaslan M, Eraslan F, Bağcı EG, Cicek N. Salicylic acid induced changes on some physiological parameters symptomatic for oxidative stress and mineral nutrition in maize (*Zea mays* L.) grown under salinity. *Journal of Plant Physiology* 2007; **164(6)**: 728-736.
- [17] Rao MV, Davis KR. Ozone-induced cell death occurs via two distinct mechanisms in Arabidopsis: the role of salicylic acid. *The Plant Journal* 1999; **17(6)**: 603-614.
- [18] Senaratna T, Touchell D, Bunn E, Dixon K. Acetyl salicylic acid (Aspirin) and salicylic acid induce multiple stress tolerance in bean and tomato plants. *Plant Growth Regulation* 2000; **30(2)**: 157-161.
- [19] Yalpani N, Silverman P, Wilson T, Kleier DA, Raskin I. Salicylic acid is a systemic signal and an inducer of pathogenesis-related proteins in virus-infected tobacco. *The Plant Cell* 1991; **3(8)**: 809-818.
- [20] Stevens J, Senaratna T, Sivasithamparam K. Salicylic acid induces salinity tolerance in tomato (*Lycopersicon esculentum* cv. Roma): associated changes in gas exchange, water relations and membrane stabilisation. *Plant Growth Regulation* 2006; **49(1)**: 77-83.
- [21] Herrera-Vásquez A, Salinas P, Holuigue L. Salicylic acid and reactive oxygen species interplay in the transcriptional control of defense genes expression. *Frontiers in Plant Science* 2015; **6**: 171.
- [22] Ahmed H, Mimouni H, Manaa A, Zid E. Salicylic acid improves the tolerance of cultivated tomato (*Solanum lycopersicum*) to salt stress. *Acta Botanica Gallica* 2010; **157(2)**: 361-368.
- [23] Qados AMA. Effects of salicylic acid on growth, yield and chemical contents of pepper (*Capsicum annum* L.) plants grown under salt stress conditions. *International Journal of Agriculture and Crop Sciences* 2015; **8(2)**: 107-113.
- [24] Horasan O, Furtana GB, Ellialtıođlu SS, Tipirdamaz R. Exogenously salicylic acid effects on growth and some physiological parameters of in vitro cultured salt stressed embryos of eggplant (*Solanum melongena* L.) cultivars differing in salt tolerance. *Fresenius Environmental Bulletin* 2017; **26(5)**: 3107-3117.
- [25] Elhindi K, Al-Amri S, Abdel-Salam E, Al-Suhaibani N. Effectiveness of salicylic acid in mitigating salt-induced adverse effects on different physio-biochemical attributes in sweet basil (*Ocimum basilicum* L.). *Journal of Plant Nutrition* 2017; **40(6)**: 908-919.
- [26] Anaya F, Fghire R, Wahbi S, Loutfi K. Influence of salicylic acid on seed germination of *Vicia faba* L. under salt stress. *Journal of the Saudi Society of Agricultural Sciences* 2018; **17(1)**: 1-8.
- [27] TÜİK. *Türkiye İstatistik Kurumu* veri tabanı. (accessed 21/12/2017).
- [28] Khah E. Effect of grafting on growth, performance and yield of aubergine (*Solanum melongena* L.) in the field and greenhouse. *Journal of Food, Agriculture and Environment* 2005; **3(3&4)**: 92-94.
- [29] Kıran S, Kuşvuran Ş, Özkay F, Özgün Ö, Sönmez K, Özbek H, Ellialtıođlu ŞŞ. Bazı patlıcan anaçlarının tuzluluk stresi koşullarındaki gelişmelerinin karşılaştırılması. *Tarım Bilimleri Araştırma Dergisi* 2015; **8(1)**: 20-30.
- [30] Özkorkmaz F, Yılmaz N. Farklı tuz konsantrasyonlarının fasulye (*Phaseolus vulgaris* L.) ve börülçede (*Vigna unguiculata* L.) çimlenme üzerine etkilerinin belirlenmesi. *Ordu Üniversitesi Bilim ve Teknoloji Dergisi* 2017; **7(2)**: 196-200.
- [31] Scardaci S, Shannon M, Grattan S, Eke A, Roberts S, Goldman-Smith S, Hill J. Water management practices can affect salinity in rice fields. *California Agriculture* 2002; **56(6)**: 184-188.
- [32] Sivritepe N, Sivritepe HO, Celik H, Katkat AV. Salinity responses of grafted grapevines: Effects of scion and rootstock genotypes. *Notulae Botanicae Horti Agrobotanici Cluj-Napoca* 2010; **38(3)**: 193-201.
- [33] Turhan E, Dardeniz A, Müftüođlu NM. Bazı Amerikan asma anaçlarının tuz stresine toleranslarının belirlenmesi. *Bahçe* 2014; **34(1)**: 11-20.
- [34] Yin C, Wang X, Duan B, Luo J, Li C. Early growth, dry matter allocation and water use efficiency of two sympatric *Populus* species as affected by water stress. *Environmental and Experimental Botany* 2005; **53(3)**: 315-322.
- [35] Yurtseven E, Ünlükara A, Top A, Tek A. Tuzluluğun ve sulama aralığının kolzada (*Brassica napus oleifera*) verime ve gelişmeye etkisi. I. *Ulusal Sulama Kongresi Bildirileri Kitabı* 2001; 8-11.
- [36] Khan W, Prithiviraj B, Smith DL. Photosynthetic responses of corn and soybean to foliar application of

- salicylates. *Journal of Plant Physiology* 2003; **160**(5): 485-492.
- [37] Kök D. Farklı salisilik asit dozlarının asma anaçlarının tuzluluğa dayanımı üzerine etkileri. *JOTAF/Tekirdağ Ziraat Fakültesi Dergisi* 2012; **9**(2): 32-40.
- [38] Shakirova FM, Sakhabutdinova AR, Bezrukova MV, Fatkhutdinova RA, Fatkhutdinova DR. Changes in the hormonal status of wheat seedlings induced by salicylic acid and salinity. *Plant Science* 2003; **164**(3): 317-322.
- [39] Karlıdag H, Yildirim E, Turan M. Salicylic acid ameliorates the adverse effect of salt stress on strawberry. *Scientia Agricola* 2009; **66**(2): 180-187.
- [40] Sarita T, Pragati M, John S, Shukla P, Ramteke P. Effect of exogenous application of salicylic acid on some morphological parameters in salt stressed tomato varieties. *Advance Research Journal of Crop Improvement* 2016; **7**(1): 100-105.
- [41] Basu R, Bose T, Roy B, Mukhopadhyay A. Auxin synergists in rooting of cuttings. *Physiologia Plantarum* 1969; **22**(4): 649-652.
- [42] Gutiérrez-Coronado MA, Trejo-López C, Larqué-Saavedra A. Effects of salicylic acid on the growth of roots and shoots in soybean. *Plant Physiology and Biochemistry* 1998; **36**(8): 563-565.
- [43] Larqué-Saavedra A, Wilkins H, Wain R. Promotion of cress root elongation in white light by 3, 5-diiodo-4-hydroxybenzoic acid. *Planta* 1975; **126**(3): 269-272.
- [44] Merwad A. Effect of potassium fertilisation and salicylic acid on yield, quality and nutrient uptake of sugar beet (*Beta vulgaris* L.) grown in saline soil. *Malaysian Journal of Soil Science* 2015; **19**: 95-105.
- [45] Daşgan HY, Koç S, Ekici B, Aktaş H, Abak K. Bazı fasulye ve börülce genotiplerinin tuz stresine tepkileri. *Alatarım* 2006; **5**(1): 23-31.
- [46] Salem A, Abdel-Aal Y, Abdel-Mohsen M, Yasin W. Tolerance of Flame Seedless grapes on own root and grafts to irrigation with saline solutions. *J. Hort. Sci. Orn. Plants* 2011; **3**: 207-219.
- [47] Grattan S, Grieve, C. Mineral element acquisition and growth response of plants grown in saline environments. *Agriculture, Ecosystems & Environment* 1992; **38**(4): 275-300.
- [48] Nable RO, Cartwright B, Lance RCM. Genotypic differences in boron accumulation in barley: Relative susceptibilities to boron deficiency and toxicity. In: *Genetic Aspects of Plant Mineral Nutrition*, El Bassam N, Dambroth M, Loughman BC. Eds. Kluwer Academic Publishers Dordrecht, The Netherlands: 1990: 243-251.
- [49] Masood S, Wimmer M, Witzel K, Zörb C, Mühling K. Interactive effects of high boron and NaCl stresses on subcellular localization of chloride and boron in wheat leaves. *Journal of Agronomy and Crop Science* 2012; **198**(3): 227-235.
- [50] Ahmed HB, Mimouni H, Manaa A, Zid E. L'acide salicylique améliore la tolérance de la tomate cultivée (*Solanum lycopersicum*) à la contrainte saline. *Acta Botanica Gallica* 2010; **157**(2): 361-368.
- [51] Cakmak I. Tansley Review No. 111. Possible roles of zinc in protecting plant cells from damage by reactive oxygen species. *The New Phytologist* 2000; **146**(2): 185-205.
- [52] Welch RM, Webb MJ, Loneragan JF. Zinc in membrane function and its role in phosphorus toxicity. In: *Proceedings of the 9th International Plant Nutrition Colloquium*, Scaife A, Ed. 1982:710-715.
- [53] Parr A, Loughman B. Boron and membrane function in plants. In: *Metals and micronutrients: Uptake and utilization by plants*, Robb DA, Pierpoint WS, eds. 1983; pp 341.
- [54] Al-Hakim, A, Hamada A. Counteraction of salinity stress on wheat plants by grain soaking in ascorbic acid, thiamin or sodium salicylate. *Biologia Plantarum* 2001; **44**(2): 253-261.
- [55] Janda T, Horváth E, Szalai G, Paldi E. Role of salicylic acid in the induction of abiotic stress tolerance. In *Salicylic acid: A plant hormone*, Ahmad SH, Ed. Springer, 2007; 91-150.
- [56] Hayat Q, Hayat S, Irfan M, Ahmad A. Effect of exogenous salicylic acid under changing environment: a review. *Environmental and Experimental Botany* 2010; **68**(1): 14-25.
- [57] Ashraf M, Khanum A. Relationship between ion accumulation and growth in two spring wheat lines differing in salt tolerance at different growth stages. *Journal of Agronomy and Crop Science* 1997; **178**(1): 39-51.
- [58] Zeng L, Poss JA, Wilson C, Draz ASE, Gregorio GB, Grieve CM. Evaluation of salt tolerance in rice genotypes by physiological characters. *Euphytica* 2003; **129**(3): 281-292.
- [59] Dasgan HY, Aktas H, Abak K, Cakmak I. Determination of screening techniques to salinity tolerance in tomatoes and investigation of genotype responses. *Plant Science* 2002; **163**(4): 695-703.
- [60] Al-Karaki GN. Growth, water use efficiency, and sodium and potassium acquisition by tomato cultivars grown under salt stress. *Journal of Plant Nutrition* 2000; **23**(1): 1-8.
- [61] Botella M, Martinez V, Pardines J, Cerda A. Salinity induced potassium deficiency in maize plants. *Journal of Plant Physiology* 1997; **150**(1-2): 200-205.
- [62] Tufail, A., Arfan, M., Gurmani, A. R., Khan, A., Bano, A., Salicylic acid induced salinity tolerance in maize (*Zea mays*). *Pak. J. Bot* 2013, 45 (S1), 75-82.